

A Peptide-mediated Fenton Reaction in Wood-degrading Fungi

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Abstract: Gt factor is a low-molecular-weight peptide isolated from the extracellular culture of wood-degrading fungus *Gloeophyllum trabeum*. It is capable of enhancing degradation of cellulose. Its action mechanism was investigated and it was found that Gt factor could reduce Fe³⁺ to Fe²⁺. Electron paramagnetic resonance (EPR) spectroscopy revealed in the presence of O₂, Gt factor could drive the formation of H₂O₂ via a superoxide anion O₂⁻ intermediate and mediate the generation of hydroxyl radical HO• in a Fenton-type reaction. All these provided evidence for the formation of HO• in some wood-degrading fungi.

Keywords: Cellulose degradation, hydroxyl radical HO•; Gt factor, electron paramagnetic resonance.

Cellulose is the most abundant resource of regenerative biomass on earth. But due to the complexity of its structure, its degradation mechanism was failed to fully understand. It makes a great limit to its full utility. In nature, wood decay is mainly accomplished by some wood-degrading fungi such as *G.trabeum et al.*. It is well-known that cellulases are the most important components of degrading cellulose. But because the cellulase activity is low, the complete utility of cellulose by enzymatic method is unsatisfactory. In 1965, Halliwell found that the hydroxyl radical HO• produced by Fenton's reagent, is quite destructive to cellulose¹.



Thus, HO• has been proposed to be involved in cellulose degradation²⁻⁶ and oxidative degradation has become a potential way in utility of wood sources. According to this proposition, low molecular-weight biochemical agent was believed to be able to penetrate plant cell wall, mediate the formation of HO• which caused cellulose decomposition⁷⁻⁸. So far lots of study involved HO• in cellulose degradation, but the methods for detecting HO• was not accurate; besides, the generation of HO• mediated by Gt factor is still not clear. In this paper, we have applied EPR spin trapping to study HO• generation mediated by Gt factor (purified peptide isolated from extracellular culture of *G.trabeum*)⁹⁻¹¹ and elucidated the pathway of HO• formation.

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Methods

The EPR spin trapping studies were performed using 5,5-dimethylpyrroline-1-oxide (DMPO) at a final concentration of 1 mmol/L. EPR spectra were recorded at room temperature using Bruker Esp 300E spectrometer. The spectrometer settings were as follows: modulation frequency 100 kHz; modulation amplitude 2.024 G; microwave power 10 MW; microwave frequency 9.8 GHz; center field 3483 G; sweeping time 1 min.

Results and Discussion

Ferrozine can react with Fe^{2+} , forming stable red compound which has characteristic absorbance at 562 nm¹². When ferrozine was added to a solution containing Gt factor and Fe^{3+} , the absorbance of the solution at 562 nm continually intensified, indicating Gt factor strongly reduced Fe^{3+} to Fe^{2+} .

DMPO was used to measure HO^\bullet generation by Gt factor in EPR study. Gt factor (5 μg) was placed in 50 mmol/L acetate buffer (pH 4.5) containing 2 mmol/L cellobiose and 0.1 mmol/L Fe^{3+} . After addition of DMPO for 30 s, EPR spectrum showed a 1: 2: 2: 1 quartet (**Figure 1**) originating from the hydroxyl radical adduct of DMPO- OH ($\alpha_N = \alpha_M = 14.9$ G). This verified the generation of HO^\bullet . Addition of EDTA (100 mmol/L) diminished HO^\bullet generation, indicating the presence of Fe^{3+} is necessary in HO^\bullet generation. When catalase was added, HO^\bullet generation decreased, indicating HO^\bullet was generated from H_2O_2 . It was further verified by the fact that after 1 mmol/L H_2O_2 was added, EPR signal of DMPO-OH significantly intensified (**Figure 2**). Thus, we can deduce that HO^\bullet formation may occur in a Fenton-type reaction. But the question is that where H_2O_2 comes from? In **Figure 3**, EPR signal of DMPO-OH was firstly produced in the presence of cellobiose, Gt factor, Fe^{3+} and O_2 . Then additional Gt factor was added, DMPO-OH signal was intensified, this is because Gt factor reduced Fe^{3+} to Fe^{2+} , which subsequently made HO^\bullet generation cycling possible. Besides, a characteristic DMPO- $\text{O}_2^{\cdot-}$ signal appeared after addition of Gt factor, suggesting the existence of superoxide anion $\text{O}_2^{\cdot-}$ in reaction system and $\text{O}_2^{\cdot-}$ formation may be driven by reduction of Fe^{3+} by Gt factor as $\text{Fe}^{2+} + \text{O}_2 \rightarrow \text{Fe}^{3+} + \text{O}_2^{\cdot-}$. This might somewhat enlighten on the mechanism of H_2O_2 generation. As we know H_2O_2 can potentially form by a one-electron transfer to O_2 to generate superoxide anion $\text{O}_2^{\cdot-}$. **Figure 3** showed the $\text{O}_2^{\cdot-}$ signal. We can conclude that H_2O_2 was generated by Gt factor *via* a superoxide anion intermediate- $\text{O}_2^{\cdot-}$. In this reaction, Gt factor acted as a reductant of Fe^{3+} , Fe^{2+} drove the formation of $\text{O}_2^{\cdot-}$ and further lead to the H_2O_2 generation.

In summary, Gt factor, a peptide from the extracellular culture of wood-degrading fungus- *G.trabeum*, could reduce Fe^{3+} to Fe^{2+} , mediate H_2O_2 formation *via* a superoxide anion intermediate - $\text{O}_2^{\cdot-}$, and lead to HO^\bullet production in a Fenton-type reaction. HO^\bullet is strongly oxidative, it could abstract electron from hydroxyl group on glucose monomer of cellulose, left a carbonyl in place of hydroxyl group, thus causing disturbance of

interchain and intrachain hydrogen bonds and then cleavage of a polysaccharide chain of cellulose¹³. FT-IR spectroscopy and X-ray diffraction analysis of dewaxed cotton fiber proved that Gt factor really could disrupt hydrogen bonds and decrease the cellulose crystallinity. The above EPR study provided evidence for HO[•] formation in some wood-degrading fungi; Furthermore, the free radical generating system described above has potential application in variety of industrial processing and environment control.

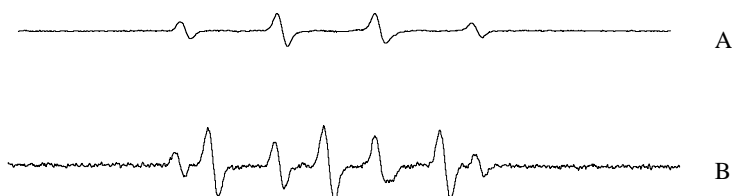
Figure 1 EPR spectrum of DMPO-OH adduct formed in the presence of Gt factor, cellobiose, Fe³⁺ and O₂.



Figure 2 EPR spectra of DMPO-OH adduct obtained from Gt factor, cellobiose, Fe³⁺ and O₂. (A) without H₂O₂; (B) 1 mmol/L H₂O₂ was added.



Figure 3 EPR spectra of DMPO-OH adduct obtained from Gt factor, cellobiose, Fe³⁺ and O₂. (A) spectrum from Gt factor, cellobiose, Fe³⁺ and O₂; (B) After addition of additional Gt factor.



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